Microscopy Core Facility

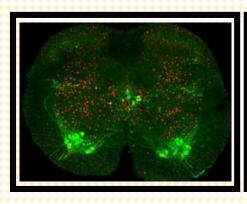
Neuroscience Engineering Collaboration Building

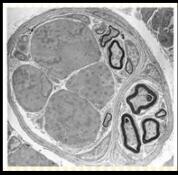
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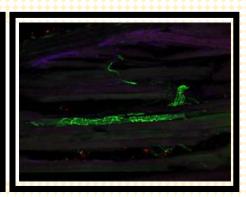
Core Personnel

Hanna Gabriel Core Manager (937) 775-3019

David Ladle Core Director (937) 775-4692







Microscopy Core Facility Equipment

Olympus FV1000

- Six excitation lines (405, 488, 458, 511, 568, 633)
- Four fluorescence detection lines (two spectral units and two based on optical filters)
- One detector for transmitted bright field (New in October 2006)

Olympus FV300

- Two excitation lines (488, 568)
- Two detectors that can be switched for different combinations of dual fluorescence or fluorescence and transmitted light simultaneous detection

Olympus 2-Photon

- Two-photon excitation is obtained with a MaiTai pulsed laser (range 700-1040 nm)
- Single photon excitation is also possible (488, 458, 511, 559, 633 lines).
- Detection can be accomplished through four non-descan detectors, three confocal detectors and one transmitted light detector
- New in September 2009

Electron Microscope

- The phillips 208S is a 100kv transmission electron microscope with excellent contrast and resolution with properly prepared specimens. The instrument, besides producing data-rich film output, is also coupled to AMT xr611 camera that permits high-resolution digital imaging
- The core provides technical help for the use of the microscope (inserting and withdrawing specimens, processing any EM negatives). Thus, usage of this instrument is always implemented with assistance by authorized core personnel.

Olympus Epi Fluorescence Spot Scope with RT Color Camera

Fluorescent Dissecting Scope with BW/Color Camera

- The Olympus Mvx10 stereomicroscope provides fluorescent view and imaging at the macro level. The optics are listed below:
- 0.63x MVX Plan Apochromat Lens has a high numerical aperture of 0.15
- 1x MVX Plan Apochromat Lens has a numerical aperture of 0.25
- 2x MVX Plan Apochromat Lens A unique and specially designed high-numerical aperture lens of 0.50 NA that features a correction-collar for correcting aberrations when imaging through plastic vessels or up to 5mm of water. This 2x PF (part of the parfocal series) objective provides superior resolution of over 1500 lines/mm

Review Computer 1

This review station has the following software:

- Image Pro Premier 9.1
- Olympus Fluoview 10 (Full Version)
- Sigma State/Plot v9
- CorelDRAW Graphics SuiteX6
- Microsoft Office 2013
- Adobe Acrobat Professional 11
- Matlab 2015

- SPSS
- Compustat
- Adobe Creative Cloud-Design Suite
- Statistica
- Spike v2
- Clampfit v5.13
- Spot Advance V9

Review Computer 2

This review stations has the following software:

- Image Pro Premier 9.1
- Olympus Fluoview 10 (Full Version)
- Adobe Acrobat Professional 11
- Matlab 2015
- SPSS
- Compustat

- Adobe Creative Cloud-Design Suite
- ClampFit v5.13
- Axon pCLAMP 10.2
- CorelDRAW Graphics SuiteX6
- Microsoft Office 2013

Review Computer 3

This review station has the following software:

- Image Pro 5.1
- Olympus Fluoview 10 (Full Version)
- CorelDRAW Graphics SuiteX6
- Adobe Acrobat Professional 11
- Matlab 2015

- SPSS
- Compustat
- Adobe Creative Cloud-Design Suite
- Spot Advance v9
- Microsoft Office 2013

HM550 Cryostat

Vibratome

Ultramictrotome MT6000

Ultramicrotome MT5000

Freezing Sliding Microtome

Stereo Investigator/ Neurolucida/ Huygens (Deconvolution)

Core Training

Request training on any equipment using iLab:

My.iLabsolutions.com

Only authorized and trained Users are allowed to use equipment in the Core Facility

Trained Users are permitted to utilize equipment during normal operating hours:

Monday – Friday 9:00 am – 5:00 pm

3-Step Process:

Initial Training –
 Meet 1:1 with Core
 Personnel to gain basic
 understanding of theory and
 techniques necessary to
 properly operate equipment
 safely and effectively.

2. 1st Supervised Session – Learn basic troubleshooting and how to optimize your results

3. 2nd Supervised Session –
It's your turn to take the lead!
Expand upon your knowledge of operating your equipment of interest, while supervised by Core Personnel

Earn 24 hour Access:

Every User has the opportunity to qualify for 24 hour access to the Microscopy Core Facility

- ☑ Log at least 20 hours of supervised usage during normal operating hours
- ☑ Demonstrate understanding of basic troubleshooting and safety procedures
- ☑ Good track record of following rules and regulations of MCF

Normal scheduling rules apply. Approval for 24 hour access is at the sole discretion of Core Personnel

Microscopy Core Facility Scheduling Guidelines:

First come, first serve scheduling starts 12am Sunday

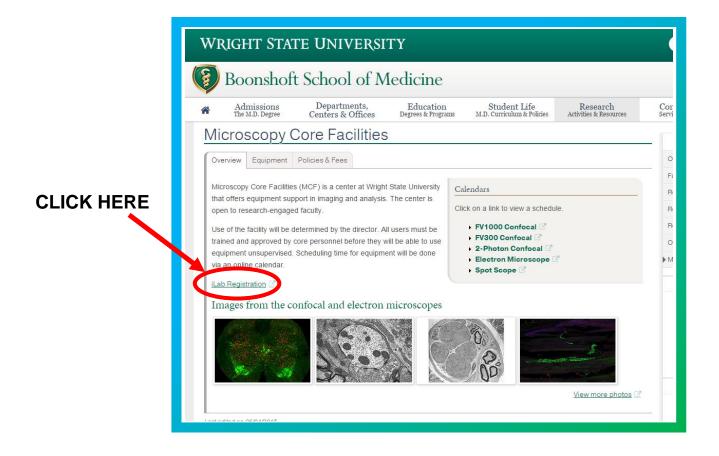
- 1. Time Slots are 4-hour increments or less
- 2. Each laboratory can reserve a maximum of two 4-hour time slots each coming week up to 2 full weeks in advance (Sunday marks the start of each week)
- Starting every Sunday at 12:00 am, available time remaining on schedule of corresponding week becomes first come, first serve
- 4. Delays in using the equipment over 15 min of scheduled time without notification to the core manager, may result in loss of scheduled time at discretion of Core Personnel
- If you are unable to use your reserved time, remove your reservation from iLab or contact Core Manager ASAP
- 6. Be PROACTIVE about scheduling! Please let the Core Manager (Hanna) know if you are having trouble getting time on the equipment Hanna.Gabriel@wright.edu; (937) 775-3019
- 7. Those individuals with 24hr access to the MCF will gain an additional allowance of up to 8 hours of advanced scheduling after normal operating hours.
 Standard open scheduling rules still apply.

Sunday	Monday	Tuesday	Wednesday	Thursday	Friday	Saturday
1	2	3	4 TODAY	5	6	7
8	9	10	11	12	13	14
Open Scheduling		4 hours		4 hours		
15	16	17	18	19	20	21
Open Scheduling			4 hours		4 hours	
22	23	24	25	26	27	28
29	30	31				

Microscopy Core Facility Register for iLab

Step 1: Visit the Microscopy Core Website at medicine.wright.edu/mcf

Step 2: Click on iLab Registration link

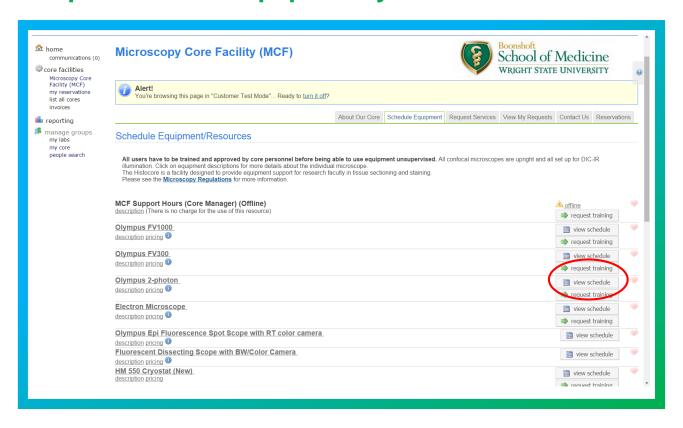


Step 3: Select your lab (PI) and follow the prompts to complete your registration

Microscopy Core Facility Scheduling/Request training on iLab

Step 1: Log into your iLab account www.my.ilabsolutions.com

Step 2: Find the equipment you want to reserve



Step 3:

Select view schedule to schedule time on equipment

Select request training to request training on equipment

Microscopy Core Facility Guidelines

Only authorized and trained Users are allowed to use the Equipment in the Core Facility

Normal operating hours: 9:00 am - 5:00 pm, Monday - Friday

Please save to guest or predestinated drive

You can remove your files from the designated computer in the histology core. Please keep in mind that the guest drive is NOT backed up and should be treated as a transitional storage space from MCF computers to each labs individual storage system

- The MCF is not responsible for data maintenance and/or data loss
- Please do not use USBs in any of the Microscopy core Computers
 This helps protect our computers from corruption
- Do not save anything on the C drive of imaging computers. This includes the desktop!
- ❖ All equipment must be cleaned and returned to proper positions with all personal belongings and trash removed or disposed.
- Scheduling rules must be followed at all times

PLEASE REPORT ANY PROBLEMS IMMEDIATELY TO CORE PERSONNEL

Core Personnel

Hanna Gabriel Core Manager (937) 775-3019 David Ladle Core Director (937) 775-4692

Microscopy Core Facility 2015 User Fees

Equipment	Rate
FV1000 (4 lasers: 405, 488, 568, 647)	\$15/hour
FV300 (2 lasers: 488, 568)	\$10/hour
2-Photon	\$20/hour
Electron Microscope – TEM	\$20/hour
Microscope Monthly Usage Fee (Includes Florescent Upright & Fluorescent Dissecting Scope)	\$10/Month
Histocore Monthly Usage Fee (Includes Cryostats, Ultramicrotomes, & Perfusion Lab)	\$10/Month
Review Station Monthly Fee (Fee is charged per user)	\$10/Month
Neurolucida/ Steroinvestigator/ Huygens (Deconvolution)	\$20/Month